MICROBIAL OXIDATION IN SYNTHESIS: PREPARATION OF 6-DEOXY CYCLITOL ANALOGUES OF *MYO*-INOSITOL 1,4,5-TRISPHOSPHATE FROM BENZENE

Steven V. Ley*, Margarita Parra, Alison J. Redgrave, Francine Sternfeld and Angel Vidal. Department of Chemistry, Imperial College of Science, Technology and Medicine, London SW7 2AY, UK.

Summary: The novel 6-deoxy, 6-deoxy-6-fluoro and 6-deoxy-6-methyl myo-inositol 1,4,5-trisphosphate derivatives (4), (5) and (6) were derived from benzene via microbial oxidation to cis-1,2-dihydroxycyclohexa-3,5-diene (2) and conversion through to the key epoxyacetonide (7).

There is currently considerable interest in the synthesis of cyclitol compounds and particularly in systems related to the cellular second messenger inositol 1,4,5-trisphosphate (IP₃) (1). ^{1,2b} Since *myo*-inositol is cheap and readily available most syntheses of inositol phosphates to date have begun with the parent cyclitol and have proceeded through a multistep protection - deprotection sequence to set up the desired pattern of substitution. We have already described a conceptually different strategy² starting from benzene and making use of a microbial oxidation using *Pseudomonas putida*^{2,3} to afford the cyclohexadiene diol (2). This approach is especially well suited to the preparation of a range of novel analogues^{2b} modified at sites other than the phosphate group, in contrast to previously reported approaches which are not readily amenable for such analogue preparation.⁴ Biological evaluation of these compounds will hopefully lead to a more detailed understanding of the phosphatidylinositol (PI) cycle regulating cellular functions as diverse as secretion, proliferation and smooth muscle contraction. We have previously published routes to IP₃ (1) and 6-*O*-methyl IP₃ (3) from (2)^{2b} and here we report the syntheses of other novel IP₃ derivatives (4)-(6).



In all of these syntheses use is made of the epoxy acetonide (7) which in turn is readily derived from (2). For the preparation of the 6-deoxy compound (4) the epoxide (7) was reacted with lithium aluminium hydride in boiling ether to afford the alcohol (8) (76%), accompanied by a small amount of the alternative ring opened product



(i) LiAlH₄, Et₂O, reflux, 2h, 88%; (ii) a. H₂, 10% Pd-C, EtOH, 39h; b. ⁿBuLi, ⁱPr₂NH, THF, tetrabenzylpyrophosphate, 56% overall; (iii) a. TMSBr, CH₂Cl₂, RT, 65 min; b. H₂O, 80 min, 79% overall.



(i) TASF, THF, reflux, 4 days, 74%; (ii) a. H₂, 10% Pd-C, EtOH, 18h; b. ⁿBuLi, ⁱPr₂NH, THF, tetrabenzylpyrophosphate, 77% overall; (iii) a. TMSBr, CH₂Cl₂, RT, 65 min; b. H₂O, 80 min, 93% overall.

(9) (12%). Debenzylation of the major product (8) by hydrogenolysis and subsequent phosphorylation with ⁿBuLi/tetrabenzylpyrophosphate gave the 1,4,5-trisphosphate (10). Total deprotection of this trisphosphate was readily achieved using Meek's TMSBr protocol.¹g Purification of the product by anion exchange HPLC (Spherisorb S5SAX column, 0.2M ammonium formate buffer at pH 4) furnished 6-deoxy IP₃ (3)⁵ in 79% overall yield from (8).

Replacement of an hydroxyl group with a fluorine atom has been considered a simple isosteric substitution in inositol synthesis,⁶ and thus analogue (5) was considered an attractive target for synthesis. Treatment of the previously described epoxy acetonide (7) with tris(dimethylamino)sulphonium difluorotrimethylsilicate (TASF)⁷ in THF at reflux afforded the fluorohydrins (11) and (12) as a 4:1 mixture readily separable by silica gel chromatography. In a similar fashion to that described above, compound (11) was hydrogenolysed and phosphorylated to afford (13). Subsequent deprotection with TMSBr and purification by anion exchange HPLC produced the novel 6-fluoro derivative of IP₃ (5) in 93% overall yield from (13).

This very versatile sequence towards a range of unique IP₃ analogue compounds was also applicable to the synthesis of the 6-deoxy-6-methyl derivative (6). Nucleophilic ring opening of (7) using lithium dimethyl(cyano)copper (I) furnished the alcohols (14) and (15) as a 21:1 mixture (76% combined yield). The alcohol (14) was then elaborated to the fully protected trisphosphate (16) in a manner analogous to that used above. Treatment with TMSBr followed by purification afforded the desired 6-deoxy-6-methyl IP₃ (6).⁹



(i) Me₂Cu(CN)Li₂, THF, -30^oC, 22h, 76%; (ii) a. H₂, 10% Pd-C, EtOH,16h; b. ⁿBuLi, ⁱPr₂NH, THF, tetrabenzylpyrophosphate, 41% overall; (iii) a. TMSBr, CH₂Cl₂, RT, 65 min; b. H₂O, 80 min, 82% overall.

The above syntheses of these IP₃ analogues further illustrates the important strategic advantage that can be gained by using microbial oxidation methodology for cyclicol preparation from aromatic substrates.

Acknowledgements

We thank the SERC and ICI plc for financial support, the Spanish Ministerio de Educacion y Ciencia for a Fellowship (to M.P.) and Dr. S.C. Taylor (ICI Bioproducts) for generous supplies of (2) and useful discussions. One of us (A.V.) acknowledges study leave from the University of Murcia, Spain.

References and Footnotes

- a) S. Ozaki, Y. Watanabe, T. Ogasawara, Y. Kondo, N. Shiotani, H. Niskii, and T. Matsuki, *Tetrahedron Lett.*, **1986**, <u>27</u>, 3157; b) A.M. Cooke, B.V. L. Potter, and R. Gigg, *Tetrahedron Lett.*, **1987**, <u>28</u>, 2305; c) C.B. Reese and J.G. Ward, *Tetrahedron Lett.*, **1987**, <u>28</u>, 2309; d) J. Gigg, R. Gigg, S. Payne and R. Conant, J. Chem. Soc., Perkin Trans. 1, **1987**, 423; e) J.P. Vacca, S.J. de Solms, and J.R. Huff, J. Am. Chem. Soc., **1987**, <u>109</u>, 3478; f) K-L. Yu and B. Fraser-Reid, *Tetrahedron Lett.*, **1988**, <u>29</u>, 979; g) J. L. Meek, F. Davidson, and F.W. Hobbs, J. Am. Chem. Soc., **1988**, <u>110</u>, 2317; h) Y-C. Liu and C-S. Chen, *Tetrahedron Lett.*, **1989**, <u>30</u>, 1617.
- a) S.V. Ley, F. Sternfeld, and S. Taylor, *Tetrahedron Lett.*, 1987, <u>28</u>, 225; b) S.V. Ley and F. Sternfeld, *Tetrahedron Lett.*, 1988, <u>29</u>, 5305.
- D.T. Gibson, J.R. Koch, and R.E. Kallio, *Biochem.*, 1968, 7, 2653; b) S.C. Taylor in "Enzymes in Organic Synthesis", Eds. R. Porter and S. Clark, CIBA Foundation Symposium III, Pitman (London), 1985; c) P.W. Howard, G.R. Stephenson, and S.C. Taylor, *J. Organomet. Chem.*, 1988, 339, C5; d) I. C. Cotterill, S.M. Roberts and J.O. Williams, *J. Chem. Soc., Chem. Commun.*, 1988, 1628; e) H.A.J. Carless and O.Z. Oak, *Tetrahedron Lett.*, 1989, 30, 1719; f) S.V. Ley and F. Sternfeld, *Tetrahedron* accepted for publication.
- a) A.M. Cooke, R. Gigg and B.V.L. Potter, J. Chem. Soc., Chem. Commun., 1987, 1525; b)
 A.M.Cooke, N.J. Noble, S. Payne, R. Gigg and B.V.L. Potter, J. Chem. Soc., Chem. Commun., 1989, 269; c) M.F. Boehm and G.D. Prestwich, Tetrahedron Lett., 1988, 29, 5217.
- 5. ¹H-NMR δ (500 MHz, D₂O, pH 9.4; <u>H</u>OD = 4.63) 4.01 (1H, s, 2-H), 3.91 (2H, br s, (C<u>H</u>OP(O)(OH)₂)₂), 3.76 (1H, br s, C<u>H</u>OP(O)(OH)₂), 3.47 (1H, d, J 6.9 Hz, 3-H), 2.17 (1H, d, J 10.5 Hz, 6 eq-H), 1.74 (1H, q, J 10.5 Hz, 6 ax-H); ³¹P-NMR δ (101 MHz, D₂O, pH 9.0) 5.93 (3P, br s).
- 6. A.P. Kozikowski, Y. Xia, and J.M. Rusnak, J. Chem. Soc., Chem. Commun., 1988, 1330.
- 7. P.J. Card and W.D. Hitz, J. Am. Chem. Soc., 1984, 106, 5348.
- 8. ¹H-NMR δ (500 MHz, D₂O, pH 9.0; <u>H</u>OD = 4.63) 4.44 (1H, dt, J 52.1 and 7.7 Hz, 6-H), 4.09 (1H, br s, 2-H), 3.99-3.95 (3H, m, (C<u>H</u>OP(O)(OH)₂)₃), 3.59 (1H, d, J 5.8 Hz, 3-H); ³¹P-NMR δ (101 MHz, D₂O, pH 9.0), 6.85 (1P, br s), 5.86 (1P, s), 5.27 (1P, br s); ¹⁹F-NMR δ (84 MHz, D₂O, pH 9.0) -196.4 (d, J 53.7Hz).
- 9. ¹H-NMR δ (500 MHz, D₂O, pH 9.0; <u>H</u>OD = 4.63) 4.09 (1H, s, 2-H), 3.97 (1H, br s, C<u>H</u>OP(O)(OH)₂), 3.59 (1H, br s, C<u>H</u>O(P)(O)(OH)₂), 3.52 (1H, dd, J 8.8 and 9.1 Hz, C<u>H</u>O(P)(O)(OH)₂), 3.47 (1H, br s, 3-H), 1.81 (1H, br s, 6-H), 0.97 (3H, d, J 4.9 Hz, Me); ³¹P-NMR δ (101 MHz, D₂O, pH 9.0), 6.74 (1P, br s), 5.47 (1P, br s), 3.90 (1P, br s).

(Received in UK 4 May 1989)